



Review

Amyotrophic lateral sclerosis as a complex genetic disease

Claire L. Simpson, Ammar Al-Chalabi *

MRC Centre for Neurodegeneration Research P 043, King's College London, Institute of Psychiatry, London SE5 8AF, UK

Received 14 May 2006; received in revised form 25 July 2006; accepted 2 August 2006

Available online 5 August 2006

Abstract

In complex diseases like ALS, there are multiple genetic and environmental factors all contributing to disease liability. The genetic factors causing susceptibility to developing ALS can be considered a spectrum from single genes with large effect sizes causing classical Mendelian ALS, to genes of smaller effect, producing apparently sporadic disease. We examine the statistical genetic principles that underpin this model and review what is known about ALS as a disease with complex genetics.

© 2006 Published by Elsevier B.V.

Keywords: Complex disease; Complex genetic; Amyotrophic lateral sclerosis; Sporadic; Association study; Gene; Genetic**1. Introduction***1.1. What is a complex disease?*

Most cases of amyotrophic lateral sclerosis (ALS) are isolated incidents. When, in 1955, Kurland and Mulder first provided evidence that in about 10% of cases there was a family history [1], the natural assumption was that the remaining sporadic cases did not have a genetic cause. This has to be viewed on the historical background of the time, when the idea that ALS might have any genetic component was surprising, ideas of Mendelian inheritance of disease were mainstream, and there was no adequate genetic model for non-Mendelian familial clustering or sporadic disease. We now have a more detailed and sophisticated understanding of the behaviour of genes, and it is reasonable to assume that in most if not all cases, there is a genetic contribution.

The discovery of mutations in SOD1 in 20% of familial cases [2,3] and between 2 and 7% of sporadic cases [4–8], indicates that at least some sporadic cases of ALS are genetic. Further evidence comes from a UK twin study, which examined mono- and dizygotic twins for concordance, having first eliminated probands from families already identified with dominant ALS

inheritance. Heritability was defined as $h^2 = 2 \times [r_{mz} - r_{dz}]$, where r_{mz} and r_{dz} are the liability correlations for mono- and dizygotic twins ranging from 0 (fully non-genetic) to 1 (fully heritable). Based on these calculations, the heritability of ALS was shown in this study to be between 38 and 85% [9]. The authors concluded that “...the finding that between 38% and 85% of variation in (ALS) is due to inherited factors indicates that even in so-called sporadic (ALS), with a conservative approach to analysis of the data, genetic influences are significant.”

Another way to examine the role genetic factors might play in sporadic ALS is to estimate the relative risk of ALS to siblings of affected individuals as compared with the population risk, given the symbol λ_s . Without detailed population risk data for ALS from genetic epidemiology, only imperfect guesses are possible. If it is assumed that since approximately 10% of cases have a family history, then the risk to a sibling of a sporadic case with an unknown family history is the risk of having a family history of ALS \times the risk of sharing a dominant allele = $1/10 \times 1/2 = 1/20$. The lifetime risk of developing ALS estimated from population data and death certificate data as between 1 in 400 and 1 in 1000 [8]. Therefore, a rough estimate of λ_s is between $(1/20)/(1/400) = 20$ and $(1/20)/(1/1000) = 50$. The problem with this approach is that it assumes dominant inheritance, but we know from many pedigrees that familial clustering may occur without obvious Mendelian patterns, and these are excluded from this calculation. Nevertheless, this value for λ_s does provide some information.

* Corresponding author. Fax: +44 20 7848 5190.

E-mail address: Ammar@iop.kcl.ac.uk (A. Al-Chalabi).

Current thinking on complex diseases like ALS is that there are multiple genetic and environmental factors all contributing to disease liability. This arises as a direct result of modern genetic statistical theory, which itself is based on an extended version of Mendelian genetics. In Mendel's description of genetic transmission of heritable characters, each "formative element" was responsible for a single character, and each parent had two formative elements for each character, transmitting one randomly to the next generation in gametes. He showed the transmission of formative elements for different characters was independent, and that some character traits were dominant to others, so that the recessive trait was masked in one generation if the parent had both an element for a dominant and recessive character simultaneously. When Mendel's ideas were rediscovered at the turn of the 20th century, the formative elements were renamed genes, the characters phenotypes, and it was discovered that one of his laws, that of independent transmission of elements for different characters, was not always true. Some phenotypes were linked together in "linkage groups", in which transmission of the gene for one phenotype affected the probability of transmission of a different gene for another phenotype. We now know the physical basis of linkage groups is the arrangement of genes in chromosomes, and the distortion by linkage of transmission frequencies of different characters from the expected 50:50 is used as a means of disease gene mapping.

Mendelian traits are well recognised, but it is difficult to explain continuous phenotypes, such as height, on the basis of single discrete genes. Height has a strong inherited component, and can be predicted reasonably accurately from parental heights. A particle theory of genetics needs to be able to explain this continuous trait without the discontinuities implied by discrete genes, but there is no such obstacle for a wave theory of genetics in which waves transmit the genetic information through generations. There was thus a battle in the early years of genetics between those who considered genes to be particles, and those who considered them to be waves. This is not as ridiculous as it may sound today, as it was thought to be a logical explanation for the transmission of continuous characters, and was a view held even by William Bateson, the first professor of genetics in history (who is also responsible for coining the word "genetics"). The solution came in 1918 when Fisher, who founded much of modern statistics, showed that three or more genes each contributing to a trait, but each following Mendel's laws, would result in a phenotypic distribution indistinguishable from a normal distribution, particularly if there was also an environmental contribution. This is because the alleles for each gene occur in pairs in each individual, and the phenotypic traits therefore follow the binomial distribution. The binomial distribution approximates the normal distribution as n increases, and with $n > 2$, the approximation is quite good. (For example, three loci with equally frequent alleles A and a, B and b, and C and c, in which capital alleles increase a trait, say IQ, by 5 and small case alleles decrease the trait by 5, result in IQs of 70, 80, 90, 100, 110, 120 and 130 with frequencies of 1:6:15:20:15:6:1). It was therefore possible to explain Mendelian traits and continuous traits using straightforward statistics applied to Mendel's laws.

Unfortunately, this approach still does not explain discrete traits showing familial clustering but without Mendelian inheritance. This was modelled successfully in 1981 with the so-called threshold liability model [10]. In this explanation, the liability to disease rather than the phenotype itself, is what is normally distributed (as in the three genes or more model), but disease only occurs once a critical threshold of liability is crossed. If the cumulative disease susceptibility burden is sufficient then the individual develops disease. This is classically illustrated with cleft lip and palate, which shows familial clustering but is not Mendelian. The "hidden" normally distributed phenotype, liability to cleft lip and palate, is in fact the speed and timing of fusion of the palatal plates. The exact speed at which they come together is unimportant so long as they meet before a critical developmental stage. This model transforms a continuous trait back into a binary phenotype/no phenotype discrete trait. The result predicted is familial clustering, but because of smaller family sizes and poor knowledge of extended families, cases can appear sporadic.

We now therefore have three models explaining three modes of genetic disease transmission: Mendelian—where discrete traits are inherited in a predictable fashion, the polygenic model for continuous traits, and the liability threshold model for a discrete trait with familial clustering which is not inherited in a Mendelian fashion. Other factors increasing the complexity of genetic predisposition to disease of which we are currently aware are: the multiple effects of single genes (pleiotropy), the interaction of multiple genes with each other (epistasis), the interaction of genes with environmental factors, splice variants of genes, variations in copy number, and post translational protein modification, but there will undoubtedly be others discovered in the future.

There is therefore complexity from the number and effect size of genes contributing to disease risk. There is a further layer of complexity at the level of phenotypic expression because the expression of one gene can alter the expression of another at transcription, at the level of molecular cell machinery, or based on the cell type, organ or system it is expressed in, and so a phenotype is rarely the product of a single gene, even if it seems so at first. Modifying genes may enhance or reduce a phenotype, produce novel phenotypes, or mask the effect of a disease gene, to produce an organism that resembles the wild-type. In Mendelian diseases, this may be manifested as reduced penetrance, for example, age or sex-dependent penetrance. Even when environmental factors are the same, traits which are inherited in a dominant manner on one genetic background can display recessive or co-dominant inheritance on another. Many other modifier effects are recognised, but all are examples of gene–gene or multiple gene–environment interactions. The "one gene, one trait" model of disease can therefore be regarded as too simple for any complete description of disease risk, even though it is useful for identifying genes for Mendelian diseases.

2. ALS susceptibility

The liability threshold model is now generally accepted as the best description and explanation of the behaviour of

complex diseases. Susceptibility to developing ALS can therefore be considered a spectrum from single genes with large effect sizes causing classical Mendelian ALS, to multiple genes of smaller effect, producing apparently sporadic disease. Those genetic causes of ALS so far identified and robustly replicated have all been in genes causing disease inherited in a classical Mendelian fashion, giving the appearance that all genetic ALS is familial and Mendelian. This is to some extent a self-fulfilling prophecy, since the gene-hunting techniques available at first required large pedigrees with clear inheritance patterns. What is now required is an approach to find the small effect genes that contribute together with each other and environmental factors to cause ALS. Hunting for such genes is possible using case-control association studies, but in the past these have been hampered by the large numbers of genetic markers and samples required to attain sufficient power. Until recently therefore, most studies have confined themselves to examining candidate genes selected because of structure, function or similarity to known genes. The problem is that in ALS little is known about the disease mechanism, making candidate gene selection difficult, haphazard, and the biased choice of a researcher. As a result, many studies performed to date have not found association, and only a few of those that have found association have been replicated.

For diseases such as ALS, with late onset and short survival, association mapping of disease genes in unrelated individuals is an important approach because obtaining the number of cases required for sufficient power in family-based studies is problematic. Genome-wide association studies, which use a strategy similar to linkage scans, uses large numbers of informative markers to identify association between allelic variants and disease. This is the ideal approach for diseases like ALS where the etiology remains largely unknown as it makes no *a priori* assumptions about the location of the variants of interest. However, the extent of linkage disequilibrium in unrelated samples is small and this means that very dense marker sets must be used on large sample sizes in order to generate sufficient power to detect association. This has meant that the costs and workload involved in such studies was unfeasible. To get around this problem, most studies have chosen a candidate gene approach where candidates are chosen based on hypotheses of disease causation. Unfortunately, such studies have so far been inconsistent and frequently underpowered. Here we review some of the most salient studies to date.

2.1. Apolipoprotein E

The difficulties in unravelling the genetic architecture of sporadic ALS are well illustrated by Apolipoprotein E (APOE) which has been studied both as a risk factor for ALS and as a modifier of various phenotypic aspects. APOE is one of the molecules responsible for the chaperoning of cholesterol through the bloodstream. It is polymorphic in the general population, with two SNPs forming three major haplotypes named as alleles, E2, E3 and E4 (the fourth, E1, does not tend to occur) [11]. Genetic variants of APOE are implicated in many abnormalities of blood lipids and cardiovascular disease, as

might be expected, but have also been shown to play a role in neurological disease. For example, the E4 allele has been associated with increased risk and earlier disease onset in late-onset familial and sporadic Alzheimer's disease [12], faster progression and earlier disease onset in multiple sclerosis [13] and poor prognosis after traumatic brain injury [14–18]. Similar studies in ALS have led to contradictory results. Some have shown no association between any of the APOE alleles and either susceptibility, or the phenotypes age at onset, bulbar onset or survival from symptom onset [19–22], while others have found a relationship between possession of the E4 allele and the phenotypes of bulbar onset [23,24] and shorter survival [25]. Although there have been many studies, the current opinion is that there is no effect of APOE on ALS risk or phenotype, but this cannot be certain. The problem with case-control studies is that they are prone to false positive association if cases and controls are not well matched. This is because differences between the two groups are not due to disease status alone, but also to other hidden attributes. To deal with this problem, proper matching of cases and controls is essential, and one way of doing this is to use family members [26]. The control can either be an unaffected sibling or the non-transmitted parental alleles. One such method is the transmission disequilibrium test [27], which is a joint test of linkage and association performed by examining the probability of transmission of a marker gene from parents to an affected offspring. If this probability is greater than the expectation of 0.5, it can be concluded that the marker is associated with disease. This approach has been used for example, to examine APOE and age of onset as a quantitative trait in ALS, showing that the age of onset of ALS for carriers of the E2 allele is about 3 years later than that for the other alleles [28].

The contradictory nature of the studies into APOE demonstrates the importance of large sample sizes, well matched controls and stringent *p*-values, for reproducible, believable associations. The key to association studies is replication of results, and without this, we should be skeptical about findings. We will now discuss a few other association studies in some detail, but for a full summary, the reader is referred to Table 1.

2.2. SOD1

ALS was first linked to chromosome 21 in 1991 [3] and mutations in SOD1 identified as accounting for 20% of familial cases [2] and 2–7% of sporadic cases [8]. Here we will only discuss its role as it relates to sporadic ALS.

SOD1 is an abundant cytosolic homodimeric protein, and each subunit contains copper and zinc ions in the active site. Its normal function is to reduce the superoxide radical to hydrogen peroxide, which can then be broken down by catalase, preventing oxidative damage. There are two other SOD molecules: SOD2 is a manganese SOD located in mitochondria and SOD3 is an extracellular SOD which also binds copper and zinc. Over 100 mutations in SOD1 have now been discovered, mostly heterozygous missense point mutations [29,30]. Unusually for an enzyme, all mutations are autosomal dominant except for the D90A mutation which can be dominant or

Table 1
A summary of association studies in ALS

Gene	Description	Reason for Investigation	Significance	References
ALAD	D-Aminolevulinic Acid Dehydratase	Lead exposure associated with ALS ALAD is involved in haem synthesis in erythrocytes	No association	[107,108]
ALS2	Alsin	Causes autosomal recessive juvenile ALS (ALS2).	No association	[109–113]
ANG	Angiogenin	ANG is functionally similar to VEGF	Association found in Scottish and Irish populations	[114,115]
APEX	Apurinic endonuclease	Defective DNA repair hypothesis of ALS etiology	May have small role but not major factor	[115]
APOE	Apolipoprotein E	Implicated in several other neurodegenerative disorders	Not associated with susceptibility. May be associated with age of onset, presentation and survival	[19,20,23–25]
AR	Androgen receptor	Causes Kennedy spinal and bulbar muscular atrophy.	No association	[116]
CCS	Copper chaperone for superoxide dismutase	Gene responsible for copper insertion into SOD1	No association	[117]
CNTF	Ciliary neurotrophic factor	Mice lacking CNTF develop mild, progressive motor neuron loss	Contradictory results	[47,49,50,53,118,119]
CYP2D6	Cytochrome p450, subfamily IID, polypeptide 6	Hypothesized poor metabolism of xenobiotics as risk factor	One reported association, not replicable	[120,121]
DCTN1	Dynactin	Disruption of dynein/dynactin complex produces motor neuron disease phenotype in mice	One reported association, not yet replicated	[95]
DNCH1	Dynein heavy chain	Mutations in Dnch1 result in progressive motor neuron degeneration in heterozygous mice, homozygotes also have Lewy-like inclusion bodies,	No association	[90,91,95]
EAAT2	Excitatory amino acid transporter 2	Excitotoxicity hypothesized to result in motor neuron degeneration	No association	[122–128]
HexA	Hexosaminidase A	HexA deficiency causes accumulation of ganglioside GM2 leads to neurodegeneration causing wide spectrum of neurological diseases	Occasionally causes rare ALS-like syndrome	[129]
HFE	Haemochromatosis	Oxidative stress is hypothesized to be implicated in neurodegeneration and misregulation of iron induces oxidative stress. Also, abnormal iron levels found in spinal cords of ALS patients	Contradictory results	[130,131]
LIF	Leukaemia inhibitory factor	LIF is same cytokine family as CNTF, involved in motor neuron survival	One reported association, not yet replicated	[132,133]
LOX	Lysyl oxidase	LOX is a copper containing enzyme and copper-induced cytotoxicity is a hypothetical mechanism of motor neuron degeneration	No association	[134]
MAO-B	Monoamine oxidase B	MAO-B generates free radicals and these are implicated in neuronal damage	Association with age at onset modification reported, not yet replicated	[135]
MAPT	Microtubule-associated protein tau	Microtubule-associated protein <i>tau</i> involved in other neurodegenerative diseases and Guam variant of ALS has neurofibrillary tangles containing aggregates of <i>tau</i>	Association reported but <i>p</i> values were weak	[136,137]
Mito	Mitochondrial DNA deletions	Accumulation of mitochondrial DNA mutations associated with aging development of degenerative diseases. In ALS, abnormal mitochondria are often found in spinal motor neurons	Associations reported, but all studies very small	[138–144]
NAIP	Neuronal apoptosis inhibitory protein	NAIP involved in related disease, spinal muscular atrophy	No association. Mutations in NAIP now considered specific for SMA	[99,101,102,106]
ND2	Subunit 2 of mitochondrial NADH dehydrogenase (Complex I)	Expression of ND2 found in Alzheimer's brains	No association	[145]
NEFH	Neurofilament, heavy chain	Neurofilament accumulation is a hallmark of ALS	Significant association, tail domain deletions present in ~1% ALS patients, not in controls. Findings have replicated in several studies	[78–81,133,146]
PRPH	peripherin	Transgenic mice over-expressing peripherin develop motor neuron degeneration	Few mutations found, not a common cause of ALS	[86,87]
PSEN1	presenilin-1	PSEN1 involved in apoptosis, a postulated mechanism for neuronal death	Weak association found, small study, not replicated	[147]

Table 1 (continued)

Gene	Description	Reason for Investigation	Significance	References
PVR	Poliovirus receptor	Poliovirus attacks motor neurons selectively, enteroviral nucleic acids found in spinal cord of ALS patients	Association with lower motor neuron disease found in one small study, not yet replicated	[148]
SETX	Senataxin	Mutations in SETX cause autosomal dominant juvenile ALS (ALS4)	New familial gene, not yet tested in sporadics	[149]
SMN1/2	Survival of motor neuron 1/2	Deletions and mutations of SMN genes cause spinal muscular atrophy	SMN genotypes which reduce SMN protein levels associated with sporadic ALS	[98,104,105,150]
Spastin and paraplegin	Hereditary spastic paraparesis	Mutations in spastin and paraplegin are the most common causes of hereditary spastic paraparesis	One case reported of young onset, slowly progressive upper and lower motor neuron syndrome with spastin mutation. No association with paraplegin	[151,152]
SNCG	Persyn	Persyn is a member of the synuclein family, γ -synuclein. Mutations in α -synuclein found in Parkinson's disease	No association	[153]
SOD1	Cu/Zn superoxide dismutase 1	Mutations in SOD1 account for 20% of familial ALS.	2–7% sporadic cases have mutations	[2,3,5,7,8,29]
SOD2	Manganese superoxide dismutase	SOD2 is a related protein of SOD1, found in mitochondria	No association	[154]
VAPB	Vesicle-associated membrane protein-associated protein B	Mutations in VAPB cause an autosomal dominant, slowly progressive ALS (ALS8)	New familial gene, not yet tested in sporadics	[155]
VDR	Vitamin D receptor	Lead exposure associated with ALS. Vitamin D can affect lead absorption and distribution	Not significant	[107,108]
VEGF	Vascular endothelial growth factor	Susceptibility	Association found in some populations	[59–61,156]

recessive depending upon the genetic background of the individual, and is polymorphic in parts of Scandinavia [31–33]. Mutant SOD1 enzyme activities vary, some being inactive (e.g. H46R) and others near normal (e.g. G93A). There seems to be no correlation between disease onset or duration and mutant SOD1 ability to scavenge superoxide, half-life, solubility, resistance to proteolytic degradation, aggregation potential, or affinity for copper, and the evidence is for a gain of a novel toxic function. How such a diverse range of mutations can result in a novel function producing the same relatively homogeneous phenotype remains a mystery.

2.3. Clinical features of SOD1 ALS

SOD1 ALS is clinically indistinguishable from sporadic ALS. The mean age of onset is consistently 46 to 47 years regardless of the underlying mutation, and in general, there is 90% penetrance by age 70. The mean duration for familial ALS in general is 3–6 years and there is little difference for familial ALS caused by SOD1 mutation. It is not unusual for there to be predominantly lower motor neuron signs, at least to start with. Bulbar onset is unusual and is usually associated with a later onset. There may occasionally be extra-motor symptoms including dementia, autonomic failure, supranuclear extra-ocular muscle weakness and bladder dysfunction.

2.4. Phenotype variation in SOD1 ALS

The phenotype observed in SOD1-mediated familial ALS is highly variable, with age-at-onset, severity and survival all

varying wildly, but there is some genotype–phenotype correlation for a few mutations. A4V is the most common ALS-associated SOD1 mutation in North America, accounting for 50% of all mutations [34]. Patients with this mutation have a common phenotype with rapid progression and death occurring at 1.4 years, rather than the 3–5 years which is more typical in other SOD1 mutations [35]. There are other SOD1 mutations with specific phenotype associations, such as I113T which has a much older mean age-at-onset compared with other SOD1 mutations, G37R, which is associated with a significantly earlier age-at-onset and longer survival, L38V, which is associated with earlier age-at-onset and G41D and G93C, which are associated with longer survival [35]. Most mutations do not relate to a single phenotype and the wide range of mutations in SOD1 cannot account for this variability as clinical variation can be observed in members of the same family [29,36]. This suggests that environmental and genetic factors are modifying the phenotype. Many mutations also show incomplete penetrance. The picture is therefore complicated even for the relatively simple situation of a single, large effect, autosomal dominant gene.

The D90A mutation is a case in point. Homozygous affected individuals are more frequent in the Torne Valley, a remote part of northern Sweden and Finland, whereas heterozygous affected are generally from other parts of the world, including southern Sweden, the UK, Belgium and France. The Torne Valley homozygotes have a predictable and relatively benign phenotype, with a generally ascending spastic weakness, long survival and occasional extramotor involvement. Heterozygous affected have a more variable phenotype

which is much more aggressive in line with other SOD1 mutations. The D90A mutation has been shown to have a single founder, about 500 generations ago, with the Torne Valley families becoming isolated approximately 50 generations ago [33,37]. One explanation for these phenotypic differences is that D90A is normally a dominant mutation, consistent with other SOD1 mutations, but that some protective factor tightly linked to SOD1 is found only in the Torne Valley families. Therefore, two copies of the D90A mutation would be required for disease development. Investigation of sequence variants in genes surrounding the SOD1 locus has not found any variants which can account for the differences in phenotype [38,39]. An alternative explanation is that all SOD1 mutations act in an additive fashion to increase the risk of developing ALS. Most mutations would have a large effect size, and so only a small contribution from other genetic or environmental factors would be needed in order to cause disease. This would account for the autosomal dominant inheritance and age-dependant penetrance. If the effect size for D90A was smaller than the other SOD1 mutations, but still more than 50% of the liability threshold, heterozygous individuals would only develop ALS if they also had a number of other risk factors contributing enough to the disease burden. A second copy of the mutation would also be sufficient to cause ALS. This would explain the pattern of recessive inheritance in the Torne Valley families and would also explain the existence of some affected heterozygotes with no family history of ALS. Some families living in particular regions with strong contributory factors would inherit the mutation phenotype as a dominant trait. This has been seen in a French family, which apparently had dominant D90A-mediated ALS [32], but was later shown to have a compound heterozygosity for a second SOD1 mutation [40]. This demonstrates that SOD1 mutations may themselves be part of a general contribution to disease liability.

2.5. Ciliary neurotrophic factor

Ciliary neurotrophic factor (CNTF) was first isolated from chick ciliary ganglia and shown to promote the survival of parasympathetic neurons *in vitro* [41]. It has been shown to have trophic and differentiating effects on many different neural cell types [42–45]. For example, Progressive Motor Neuropathy (*pnn*) is a naturally occurring autosomal recessive mutation found in mice which resembles spinal muscular atrophy in humans. When mice homozygous for the *pnn* mutation were treated with CNTF at symptom onset, they showed prolonged survival, improved motor function and fewer of the cellular changes associated with neurodegeneration [46]. Conversely, inactivation of the mouse CNTF gene is not lethal, but the mice develop progressive muscle atrophy, loss of motor neurons and muscle weakness [47], suggesting that in mice at least, CNTF expression is not essential for spinal motor neuron development but is necessary for postnatal neuronal survival.

These results suggest a possible role for CNTF abnormalities in human neurological and psychiatric disorders. Approximately 2% of the human population are homozygous for

mutations that inactivate CNTF. So far investigations of such mutations in unipolar affective disorder, schizophrenia and Alzheimer's disease have failed to find any association [48]. Similarly, in ALS, most formal studies of null CNTF mutations have not found association [49–51]. There is however a report of a family with ALS due to SOD1 mutation, one member of which was homozygous null for CNTF and had a much younger onset and more rapid disease course than other affected family members [52]. It was suggested that CNTF was modifying age of onset in this family. This was supported by the observation that mice transgenic for the G93A SOD1 mutation crossed with homozygous null CNTF mice had earlier disease onset than the wild type, but disease duration was unaffected. Although this is a compelling argument, the pattern of SOD1 and CNTF inheritance in this family can equally be explained by chance. A later study of 400 ALS patients found no association between null mutations in CNTF and age of onset, disease progression, survival or clinical presentation [53]. It is therefore difficult to conclude that CNTF is a major modifier of ALS phenotype without further evidence.

2.6. Vascular endothelial growth factor and angiogenin

Vascular endothelial growth factor (VEGF) regulates the growth of new blood vessels and induces vascular permeability. However, VEGF also has additional non-vascular functions. Neuronal axon pathfinding is mediated by semaphorins acting on neuropilin receptors. These receptors are also acted on by various members of the VEGF family [54,55], which implies a role for VEGF in neuronal growth and repair. Additionally, VEGF receptors have been discovered on neurons and astrocytes [56] and on the neural progenitor cells of the retina [57], which suggests that VEGF can act in a neurotrophic and neuroprotective role in the central and peripheral nervous system. Other pro-angiogenic growth factors such as basic fibroblast growth factor (bFGF) and platelet derived growth factor (PDGF) have also been shown to be neurotrophic [58]. It is perhaps not surprising therefore that transgenic mice with a deletion of the hypoxia-response element in the VEGF promoter (*Vegfa*^{Δ6}) develop a progressive motor neuron degeneration at about 5 months of age, reminiscent of ALS [59] (although this was a surprise when first discovered). These mice have reduced levels of VEGF in neural tissue, and induction of VEGF expression by hypoxia in the spinal cord and the brain is diminished.

Following the initial findings in mice, a large multi-centre study was performed to analyse the equivalent region in humans. SNPs and SNP haplotypes were associated with ALS in the Swedish population, Belgian population and a UK population from the Birmingham area. A fourth population from London in the UK did not show an association. In a meta-analysis of the separate centre studies, individuals homozygous for two haplotypes in the VEGF promoter/leader sequence were found to have 1.8 times increased risk of developing ALS [60]. This is one of the largest association studies in ALS to date and included almost 2000 people. It provides good evidence for a true association of VEGF haplotype and susceptibility to ALS,

but there have been criticisms of the statistical interpretation of the findings.

Other studies have been unable to replicate these results [61–64], which may reflect the small sizes of these studies compared to the original study. A possible weak association with affection status and sex has been found [64] and it has been hypothesized that high female to male ratios are needed to detect the association, since this study and the London population in the original study failed to show an association, and they both had low female to male ratios compared to the other populations examined. Further study will have to be done to show whether this is a real association or merely coincidence. Nevertheless, these same haplotypes are associated with a decrease in levels of circulating VEGF and reduction in gene transcription and therefore have a functional effect on the neuronal response to hypoxia. Given the findings in mice, this supports a role in humans, but further studies are required.

A related protein, also involved in the response to hypoxia is angiogenin. This protein is interesting because its gene lies close to that of APEX nuclease which has been implicated in ALS (Table 1), and it is therefore possible that the signal in APEX nuclease is in fact a result of linkage disequilibrium with a true functional variant in angiogenin (ANG). Angiogenin is functionally similar to VEGF, is an inducer of neovascularisation and is required for cell proliferation induced by other angiogenic proteins such as VEGF [65]. A common haplotype associated with a mutation in the angiogenin gene has been identified in patients of Scottish and Irish descent, although an association was not found in other populations studied [65]. Seven missense mutations were identified in both familial and sporadic ALS patients, of which five were within highly conserved residues in catalytic sites of the ANG protein, suggesting a direct effect on the activity of ANG. There is an interaction between ANG and VEGF, in that downregulation of angiogenin reduces VEGF-induced rRNA transcription, and inhibition of nuclear translocation of ANG completely knocks out the angiogenic activity of VEGF. As described above, mice lacking VEGF are known to develop motor neuron degeneration [59] and this provides a possible hypothesis of disease causation for ANG mutations. Interestingly, the effect of ANG mutations seem to be population specific, being found mainly in those with Irish/Scottish ancestry, and it will be important to discover the modifying factors that contribute to disease risk to produce this pattern.

2.7. Neurofilaments and peripherin

One of the characteristic pathological findings in ALS is the presence of accumulations of neurofilaments in the perikarya and proximal axons [66–69]. Neurofilaments are a type of intermediate filament found in neural cells and are thought to play a critical role in supporting the long thin axonal processes. Neurofilaments are classified by size as light (NEFL), medium (NEFM) and heavy (NEFH) chains, and assemble as heteropolymers with a NEFL core with NEFM and NEFH arranged around the outside. Chains are distinguished on the basis of tail length, with NEFM and NEFH containing a repeating lysine–

serine–proline (KSP) motif which can be variably phosphorylated. Tail phosphorylation causes extension and electrostatic repulsion and is thought to regulate axonal calibre and interaction with other cytoskeletal components.

Overexpression of the mouse light chain neurofilament gene, NEFL and the human heavy chain neurofilament gene, NEFH, had been shown to cause a neurodegenerative syndrome in transgenic mice [70,71] leading to the hypothesis that overexpression of neurofilaments could be an important part of human disease development. This view was supported by the evidence that disruption of the neurofilament genes to the point where neurofilaments were completely absent protected mice containing the human G85R SOD1 transgene from neurodegeneration [72]. The authors hypothesized that although neurofilaments are important for neuronal survival in development, they contribute to the vulnerability of motor neurons to the toxic effect of G85R SOD1.

When transgenic mice containing mutant SOD1 were crossed with transgenic mice overexpressing the heavy and light chain neurofilament gene [73–75], surprisingly these double transgenic mice had a large improvement in their lifespans and analysis of the spinal cord of 1 year old mice revealed that the massive neurodegeneration typically seen in mutant SOD1 mice was markedly less in double transgenic littermates [73–75]. This confusing picture of neurofilaments in SOD1-mediated ALS suggests that more studies are needed in this area.

Studies of neurofilament mutations in ALS patients have been suggestive of a primary but not major role in ALS. The first of these identified two common variants of the NEFH tail, differing in KSP repeat length, neither variant associated with susceptibility to ALS [76]. A further study by the same group found a codon deletion in five affected individuals and no controls [77]. Two family based studies did not find linkage of the region or mutation in about a hundred patients, suggesting that NEFH variants are not a cause of familial ALS [78,79]. A study of sporadic patients from Scandinavia and the UK identified several mutations in NEFH, accounting for approximately 1% of sporadic ALS cases, and this study also identified a deletion in an individual with familial ALS, although it was not possible to tell if it segregated with disease in this family [80]. Interestingly, all were deletions of multiples of whole repeat motifs, and this pattern also applied to an 84 bp insertion found in a different study [81]. Disease-associated deletions are always paired with the longer wild-type NEFH allele, and the insertion paired with the shorter wild-type NEFH allele. It is possible that the difference in tail lengths is one of the factors increasing susceptibility to ALS, but the picture remains unclear, with the latest study unable to positively confirm the association for NEFH or the other neurofilament subunits [82]. NEFL mutations have been found in a related motor neuron disease, Charcot–Marie–Tooth type 2E [83,84], a hereditary motor and sensory neuropathy.

Peripherin is another type of intermediate filament, preferentially expressed in the peripheral nervous system [85]. A frameshift mutation has been identified in one ALS patient [86], and this was not found in controls. This mutation produced a

truncated protein which when transfected into SW13 cell cultures was unable to self-assemble into neurofilament structures and disrupted NEFL and NEFM assembly into the intermediate filament network. One other mutation has been identified in a patient with sporadic ALS [87]. Immunocytochemistry revealed unusual large aggregates within the spinal motor neurons which contained both neurofilaments and peripherin. Transfected into mice, the mutant protein spontaneously formed aggregates, implying that the mutation was interfering with peripherin assembly.

These studies suggest that genetic variants of intermediate filaments may be a primary cause or contribute significantly towards susceptibility in a small proportion of cases.

2.8. Dynein and dynactin

The neurofilament network is a key part of the axonal transport system, and the pathological findings in ALS taken with the potential primary role of neurofilament mutations, suggest that impairment of axonal transport may be an important step in ALS pathogenesis. In keeping with this, SOD1 mouse models of ALS display impaired retrograde axonal transport [88]. This form of transport is mediated by dyneins, which are microtubule-activated ATPases responsible for transport towards the (–) end of microtubules. They are very large multimeric proteins composed of two or three heavy chains complexed with variable numbers of intermediate and light chains, and require a complex of microtubule-binding proteins that link the cargo to the microtubules. One such complex is dynactin, a well characterised set of at least 8 different subunits, enhancing dynein-dependent motility.

Homozygous loss of the cytoplasmic dynein heavy chain 1 gene (DNCHC1) is often lethal at an early stage of embryogenesis in mice, but heterozygotes are normal [89]. Mutagenesis of this gene has produced two mouse phenotypes with age-related progressive loss of muscle tone and motor ability. Such mutations specifically result in defective fast retrograde transport only observed in alpha motor neurons. Homozygous mice surviving to birth are severely affected and cannot feed or move, dying within 24 h of birth [90]. Lewy-like inclusion bodies and significant loss of spinal anterior horn cells can be observed in homozygotes *in utero*, and these are similar pathological findings to those observed in human ALS and other motor neuron degeneration phenotypes. Despite this, no mutations in human dynein have been found in ALS [91] although mutant SOD1 has been shown to interact with the dynein/dynactin complex [92]. The story for dynactin is however a little different. In *Drosophila*, disrupting the dynactin complex increases the frequency and extent of synaptic retraction events at the neuromuscular junction [93] and this suggests that dynactin is critical for synapse stabilisation. Transgenic mice overexpressing dynamitin, one of the dynactin complex subunits, show inhibited retrograde axonal transport, caused by disruption of the dynein–dynactin complex [94]. These mice develop a late-onset, slowly progressing neurodegeneration similar to human ALS. Dynactin mutations have now been identified in a few familial and sporadic ALS patients

[95–97] and it seems likely that genetic variants which impair axonal transport increase susceptibility to ALS.

2.9. SMN genes and copy number variation in complex diseases

Childhood-onset spinal muscular atrophy (SMA) is a disease confined to lower motor neurons and is caused by deletions or mutations in the Survival Motor Neuron (SMN) gene. There are two copies of SMN in humans; SMA is only caused by mutations or deletions in SMN1, but SMN2 copy number is associated with modification of disease severity. Because of the similarities between SMA and ALS, various studies have examined the relationship between the SMN genes and ALS. Studies looking for mutations in SMN1 in ALS patients have not found any association [98–102]. Studies of SMN2 deletion have shown conflicting results [103,104] but these were small studies and it is therefore difficult to draw firm conclusions. Because the chromosome 5 region containing the SMN genes is duplicated, some studies have examined copy number. These have shown that copy number variation that reduces the amount of SMN protein expressed is associated with ALS susceptibility and increased disease severity [98,105]. Further evidence in support of this comes from studies of transgenic SOD1-G86R mice crossbred onto differing genetic backgrounds, which show significantly altered age at onset on some backgrounds compared to others [106]. This effect has been linked to a region of the mouse genome syntenic to human chromosome 5 and which contains the mouse SMN gene.

Both SMN1 and SMN2 contribute to total SMN protein levels. However, SMN2 is less efficient than SMN1 and produces only 20% of SMN transcripts. To assess copy number impact on total SMN protein levels, it is possible to estimate SMN protein level using a formula derived by Veldink and colleagues: SMN protein level = SMN1 copy number + 0.20 × SMN2 copy number [105]. For example, 2 copies of SMN1 and 2 copies of SMN2 would give an SMN level of 2.4. Patients with ALS have been shown to have significantly less total SMN protein than controls, the majority having an SMN level ≤2.2, whereas most controls had a level ≥2.4.

Copy number variation at the SMN locus may therefore be important in ALS, and copy number in general is being increasingly recognised as a significant cause of genetic disease.

3. Conclusion

The number of replicated association findings in ALS remains depressingly small. No obvious themes of involved proteins have yet emerged, but this remains the hope. Although we continue to make small advances in our understanding of the genetic factors which contribute to disease risk or modify disease phenotype in ALS, there is a great need for large-scale association studies. Despite many years of research and the discovery of a number of genes which cause familial ALS, these still only account for a very small proportion of cases. Candidate gene association studies are limited by their narrow focus on small regions of the

genome, and in a disease like ALS, they can only be informed by uncertain hypotheses of the biological basis of the disease, or by genes already known to cause familial ALS and related diseases. The techniques for genotyping large numbers of genetic markers in a feasible time and at a reasonable cost are now with us. Genome-wide association studies are now a realistic approach for the investigation of the genetic component to sporadic ALS. Recent studies into diseases such as age related macular degeneration, multiple sclerosis, and rheumatoid arthritis show that these sorts of association studies can find robust, replicable associations. These represent our best hope of elucidating mechanisms that underlie the disease and provide new targets for palliative, preventative and curative treatments.

References

- [1] L.T. Kurland, D.W. Mulder, Epidemiologic investigations of amyotrophic lateral sclerosis. 2. Familial aggregations indicative of dominant inheritance, *I Neurology* 5 (1955) 182–196.
- [2] D.R. Rosen, T. Siddique, D. Patterson, D.A. Figlewicz, P. Sapp, A. Hentati, D. Donaldson, J. Goto, J.P. O'Regan, H.X. Deng, et al., Mutations in Cu/Zn superoxide dismutase gene are associated with familial amyotrophic lateral sclerosis, *Nature* 362 (1993) 59–62.
- [3] T. Siddique, D.A. Figlewicz, M.A. Pericak-Vance, J.L. Haines, G. Rouleau, A.J. Jeffers, P. Sapp, W.Y. Hung, J. Bebout, D. McKenna-Yasek, et al., Linkage of a gene causing familial amyotrophic lateral sclerosis to chromosome 21 and evidence of genetic-locus heterogeneity, *N. Engl. J. Med.* 324 (1991) 1381–1384.
- [4] P.M. Andersen, P. Nilsson, V. Ala-Hurula, M.L. Keranen, I. Tarvainen, T. Haltia, L. Nilsson, M. Binzer, L. Forsgren, S.L. Marklund, Amyotrophic lateral sclerosis associated with homozygosity for an Asp90Ala mutation in CuZn-superoxide dismutase, *Nat. Genet.* 10 (1995) 61–66.
- [5] M. Jackson, A. Al-Chalabi, Z.E. Enayat, B. Chioza, P.N. Leigh, K.E. Morrison, Copper/zinc superoxide dismutase 1 and sporadic amyotrophic lateral sclerosis: analysis of 155 cases and identification of a novel insertion mutation, *Ann. Neurol.* 42 (1997) 803–807.
- [6] C.T. Jones, P.J. Shaw, G. Chari, D.J. Brock, Identification of a novel exon 4 SOD1 mutation in a sporadic amyotrophic lateral sclerosis patient, *Mol. Cell. Probes* 8 (1994) 329–330.
- [7] C.T. Jones, R.J. Swingle, D.J. Brock, Identification of a novel SOD1 mutation in an apparently sporadic amyotrophic lateral sclerosis patient and the detection of Ile113Thr in three others, *Hum. Mol. Genet.* 3 (1994) 649–650.
- [8] C.A.S. Johnston, B.R., M.R. Turner, R. Gray, H.-M. Blunt, D. Butt, M.A. Ampong, C.E. Shaw, P.N. Leigh, A. Al-Chalabi, Amyotrophic Lateral Sclerosis in an Urban Setting: a population based study of inner city London. *J. Neurol.* (in press).
- [9] A.J. Graham, A.M. Macdonald, C.H. Hawkes, British motor neuron disease twin study, *J. Neurol., Neurosurg. Psychiatry* 62 (1997) 562–569.
- [10] D.S. Falconer, *Introduction to Quantitative Genetics*, 2nd ed. Longman, London, 1981.
- [11] P. de Knijff, A.M. van den Maagdenberg, R.R. Frants, L.M. Havekes, Genetic heterogeneity of apolipoprotein E and its influence on plasma lipid and lipoprotein levels, *Hum. Mutat.* 4 (1994) 178–194.
- [12] E.H. Corder, A.M. Saunders, W.J. Strittmatter, D.E. Schmechel, P.C. Gaskell, G.W. Small, A.D. Roses, J.L. Haines, M.A. Pericak-Vance, Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families, *Science* 261 (1993) 921–923.
- [13] J. Chapman, S. Vinokurov, A. Achiron, D.M. Karussis, K. Mitosek-Szewczyk, M. Birnbaum, D.M. Michaelson, A.D. Korczyn, APOE genotype is a major predictor of long-term progression of disability in MS, *Neurology* 56 (2001) 312–316.
- [14] F.C. Crawford, R.D. Vanderploeg, M.J. Freeman, S. Singh, M. Waisman, L. Michaels, L. Abdullah, D. Warden, R. Lipsky, A. Salazar, M.J. Mullan, APOE genotype influences acquisition and recall following traumatic brain injury, *Neurology* 58 (2002) 1115–1118.
- [15] G. Friedman, P. Froom, L. Sazbon, I. Grinblatt, M. Shochina, J. Tsenter, S. Babaey, B. Yehuda, Z. Groswasser, Apolipoprotein E-epsilon4 genotype predicts a poor outcome in survivors of traumatic brain injury, *Neurology* 52 (1999) 244–248.
- [16] C.L. Lendon, J.M. Harris, A.L. Pritchard, J.A. Nicoll, G.M. Teasdale, G. Murray, Genetic variation of the APOE promoter and outcome after head injury, *Neurology* 61 (2003) 683–685.
- [17] G.M. Teasdale, G.D. Murray, J.A. Nicoll, The association between APOE epsilon4, age and outcome after head injury: a prospective cohort study, *Brain* 128 (2005) 2556–2561.
- [18] G.M. Teasdale, J.A. Nicoll, G. Murray, M. Fiddes, Association of apolipoprotein E polymorphism with outcome after head injury, *Lancet* 350 (1997) 1069–1071.
- [19] S. Mui, G.W. Rebeck, D. McKenna-Yasek, B.T. Hyman, R.H. Brown Jr., Apolipoprotein E epsilon 4 allele is not associated with earlier age at onset in amyotrophic lateral sclerosis, *Ann. Neurol.* 38 (1995) 460–463.
- [20] T. Siddique, M.A. Pericak-Vance, J. Caliendo, S.T. Hong, W.Y. Hung, J. Kaplan, D. McKenna-Yasek, J.B. Rimmer, P. Sapp, A.M. Saunders, W.K. Scott, N. Siddique, J.L. Haines, R.H. Brown, Lack of association between apolipoprotein E genotype and sporadic amyotrophic lateral sclerosis, *Neurogenetics* 1 (1998) 213–216.
- [21] R. Bachus, S. Bader, R. Gessner, A.C. Ludolph, Lack of association of apolipoprotein E epsilon 4 allele with bulbar-onset motor neuron disease, *Ann. Neurol.* 41 (1997) 417.
- [22] R.G. Smith, L.J. Haverkamp, S. Case, V. Appel, S.H. Appel, Apolipoprotein E epsilon 4 in bulbar-onset motor neuron disease, *Lancet* 348 (1996) 334–335.
- [23] A. Al-Chalabi, Z.E. Enayat, M.C. Bakker, P.C. Sham, D.M. Ball, C.E. Shaw, C.M. Lloyd, J.F. Powell, P.N. Leigh, Association of apolipoprotein E epsilon 4 allele with bulbar-onset motor neuron disease, *Lancet* 347 (1996) 159–160.
- [24] B. Moulard, A. Sefiani, A. Laamri, A. Malafosse, W. Camu, Apolipoprotein E genotyping in sporadic amyotrophic lateral sclerosis: evidence for a major influence on the clinical presentation and prognosis, *J. Neurol. Sci.* 139 (1996) 34–37 (Suppl.).
- [25] V.E. Drory, M. Birnbaum, A.D. Korczyn, J. Chapman, Association of APOE epsilon4 allele with survival in amyotrophic lateral sclerosis, *J. Neurol. Sci.* 190 (2001) 17–20.
- [26] C.a.R. Falk, P., Haplotype relative risks: an easy reliable way to construct a proper control sample for risk calculations, *Ann. Hum. Genet.* 51 (1987) 227–233.
- [27] R.a.M. Spielman, R.E., W.J. Ewens, Transmission test for linkage disequilibrium: the insulin gene region and insulin-dependent diabetes mellitus (IDDM), *Am. J. Hum. Genet.* 52 (1993) 506–516.
- [28] Y.J. Li, M.A. Pericak-Vance, J.L. Haines, N. Siddique, D. McKenna-Yasek, W.Y. Hung, P. Sapp, C.I. Allen, W. Chen, B. Hosler, A.M. Saunders, L.M. Dellefave, R.H. Brown Jr., T. Siddique, Apolipoprotein E is associated with age at onset of amyotrophic lateral sclerosis, *Neurogenetics* 5 (2004) 209–213.
- [29] P.M. Andersen, P. Nilsson, M.L. Keranen, L. Forsgren, J. Hagglund, M. Karlsborg, L.O. Ronnevi, O. Gredal, S.L. Marklund, Phenotypic heterogeneity in motor neuron disease patients with CuZn-superoxide dismutase mutations in Scandinavia, *Brain* 120 (Pt 10) (1997) 1723–1737.
- [30] P.M. Andersen, K.B. Sims, W.W. Xin, R. Kiely, G. O'Neill, J. Ravits, E. Pioro, Y. Harati, R.D. Brower, J.S. Levine, H.U. Heinicke, W. Seltzer, M. Boss, R.H. Brown Jr., Sixteen novel mutations in the Cu/Zn superoxide dismutase gene in amyotrophic lateral sclerosis: a decade of discoveries, defects and disputes, *Amyotroph. Lateral. Scler. Other Mot. Neuron Disord.* 4 (2003) 62–73.
- [31] T. Aguirre, G. Matthijs, W. Robberecht, P. Tilkin, J.J. Cassiman, Mutational analysis of the Cu/Zn superoxide dismutase gene in 23 familial and 69 sporadic cases of amyotrophic lateral sclerosis in Belgium, *Eur. J. Hum. Genet.* 7 (1999) 599–602.
- [32] W. Robberecht, T. Aguirre, L. Van den Bosch, P. Tilkin, J.J. Cassiman, G. Matthijs, D90A heterozygosity in the SOD1 gene is associated with

- familial and apparently sporadic amyotrophic lateral sclerosis, *Neurology* 47 (1996) 1336–1339.
- [33] A. Al-Chalabi, P.M. Andersen, B. Chioza, C. Shaw, P.C. Sham, W. Robberecht, G. Matthijs, W. Camu, S.L. Marklund, L. Forsgren, G. Rouleau, N.G. Laing, P.V. Hulse, T. Siddique, P.N. Leigh, J.F. Powell, Recessive amyotrophic lateral sclerosis families with the D90A SOD1 mutation share a common founder: evidence for a linked protective factor, *Hum. Mol. Genet.* 7 (1998) 2045–2050.
- [34] M.E. Cudkowicz, R.H. Brown Jr., An update on superoxide dismutase 1 in familial amyotrophic lateral sclerosis, *J. Neurol. Sci.* 139 (1996) 10–15 (Suppl.).
- [35] M.E. Cudkowicz, D. McKenna-Yasek, P.E. Sapp, W. Chin, B. Geller, D.L. Hayden, D.A. Schoenfeld, B.A. Hosler, H.R. Horvitz, R.H. Brown, Epidemiology of mutations in superoxide dismutase in amyotrophic lateral sclerosis, *Ann. Neurol.* 41 (1997) 210–221.
- [36] L. Regal, L. Vanopdenbosch, P. Tilkin, L. Van den Bosch, V. Thijs, R. Sciot, W. Robberecht, The G93C mutation in superoxide dismutase 1: clinicopathologic phenotype and prognosis, *Arch. Neurol.* 63 (2006) 262–267.
- [37] M.J. Parton, W. Broom, P.M. Andersen, A. Al-Chalabi, P. Nigel Leigh, J.F. Powell, C.E. Shaw, D90A-SOD1 mediated amyotrophic lateral sclerosis: a single founder for all cases with evidence for a Cis-acting disease modifier in the recessive haplotype, *Hum. Mutat.* 20 (2002) 473.
- [38] W.J. Broom, C. Russ, P.C. Sapp, D. McKenna-Yasek, B.A. Hosler, P.M. Andersen, R.H. Brown Jr., Variants in candidate ALS modifier genes linked to Cu/Zn superoxide dismutase do not explain divergent survival phenotypes, *Neurosci. Lett.* 392 (2006) 52–57.
- [39] P.A. Jonsson, A. Backstrand, P.M. Andersen, J. Jacobsson, M. Parton, C. Shaw, R. Swigler, P.J. Shaw, W. Robberecht, A.C. Ludolph, T. Siddique, V.I. Skvortsova, S.L. Marklund, CuZn-superoxide dismutase in D90A heterozygotes from recessive and dominant ALS pedigrees, *Neurobiol. Dis.* 10 (2002) 327–333.
- [40] C.K. Hand, V. Mayeux-Portas, J. Khoris, V. Briolotti, P. Clavelou, W. Camu, G.A. Rouleau, Compound heterozygous D90A and D96N SOD1 mutations in a recessive amyotrophic lateral sclerosis family, *Ann. Neurol.* 49 (2001) 267–271.
- [41] R. Adler, K.B. Landa, M. Manthorpe, S. Varon, Cholinergic neurotrophic factors: intraocular distribution of trophic activity for ciliary neurons, *Science* 204 (1979) 1434–1436.
- [42] A.C. Lo, L. Li, R.W. Oppenheim, D. Prevette, L.J. Houenou, Ciliary neurotrophic factor promotes the survival of spinal sensory neurons following axotomy but not during the period of programmed cell death, *Exp. Neurol.* 134 (1995) 49–55.
- [43] M. Sendtner, Y. Arakawa, K.A. Stockli, G.W. Kreutzberg, H. Thoenen, Effect of ciliary neurotrophic factor (CNTF) on motoneuron survival, *J. Cell Sci., Suppl.* 15 (1991) 103–109.
- [44] M. Sendtner, G.W. Kreutzberg, H. Thoenen, Ciliary neurotrophic factor prevents the degeneration of motor neurons after axotomy, *Nature* 345 (1990) 440–441.
- [45] N.Y. Ip, G.D. Yancopoulos, The neurotrophins and CNTF: two families of collaborative neurotrophic factors, *Annu. Rev. Neurosci.* 19 (1996) 491–515.
- [46] M. Sendtner, H. Schmalbruch, K.A. Stockli, P. Carroll, G.W. Kreutzberg, H. Thoenen, Ciliary neurotrophic factor prevents degeneration of motor neurons in mouse mutant progressive motor neuronopathy, *Nature* 358 (1992) 502–504.
- [47] Y. Masu, E. Wolf, B. Holtmann, M. Sendtner, G. Brem, H. Thoenen, Disruption of the CNTF gene results in motor neuron degeneration, *Nature* 365 (1993) 27–32.
- [48] J. Gelemtier, C. Van Dyck, D.P. van Kammen, R. Malison, L.H. Price, J.F. Cubells, R. Berman, D.S. Charney, G. Heninger, Ciliary neurotrophic factor null allele frequencies in schizophrenia, affective disorders, and Alzheimer's disease, *Am. J. Med. Genet.* 74 (1997) 497–500.
- [49] R.W. Orrell, A.W. King, R.J. Lane, J.S. de Belleruche, Investigation of a null mutation of the CNTF gene in familial amyotrophic lateral sclerosis, *J. Neurol. Sci.* 132 (1995) 126–128.
- [50] R. Takahashi, Deficiency of human ciliary neurotrophic factor (CNTF) is not causally related to amyotrophic lateral sclerosis (ALS), *Rinsho Shinkeigaku* 35 (1995) 1543–1545.
- [51] R. Takahashi, H. Yokoji, H. Misawa, M. Hayashi, J. Hu, T. Deguchi, A null mutation in the human CNTF gene is not causally related to neurological diseases, *Nat. Genet.* 7 (1994) 79–84.
- [52] R. Giess, R. Goetz, B. Schrank, G. Ochs, M. Sendtner, K. Toyka, Potential implications of a ciliary neurotrophic factor gene mutation in a German population of patients with motor neuron disease, *Muscle Nerve* 21 (1998) 236–238.
- [53] A. Al-Chalabi, M.D. Scheffler, B.N. Smith, M.J. Parton, M.E. Cudkowicz, P.M. Andersen, D.L. Hayden, V.K. Hansen, M.R. Turner, C.E. Shaw, P.N. Leigh, R.H. Brown Jr., Ciliary neurotrophic factor genotype does not influence clinical phenotype in amyotrophic lateral sclerosis, *Ann. Neurol.* 54 (2003) 130–134.
- [54] G.H. Fong, J. Rossant, M. Gertsenstein, M.L. Breitman, Role of the Flt-1 receptor tyrosine kinase in regulating the assembly of vascular endothelium, *Nature* 376 (1995) 66–70.
- [55] S. Soker, S. Takashima, H.Q. Miao, G. Neufeld, M. Klagsbrun, Neuropilin-1 is expressed by endothelial and tumor cells as an isoform-specific receptor for vascular endothelial growth factor, *Cell* 92 (1998) 735–745.
- [56] F. Lennmyr, K.A. Ata, K. Funa, Y. Olsson, A. Terent, Expression of vascular endothelial growth factor (VEGF) and its receptors (Flt-1 and Flk-1) following permanent and transient occlusion of the middle cerebral artery in the rat, *J. Neuropathol. Exp. Neurol.* 57 (1998) 874–882.
- [57] K. Yang, C.L. Cepko, Flk-1, a receptor for vascular endothelial growth factor (VEGF), is expressed by retinal progenitor cells, *J. Neurosci.* 16 (1996) 6089–6099.
- [58] M. Sondell, G. Lundborg, M. Kanje, Vascular endothelial growth factor has neurotrophic activity and stimulates axonal outgrowth, enhancing cell survival and Schwann cell proliferation in the peripheral nervous system, *J. Neurosci.* 19 (1999) 5731–5740.
- [59] B. Oosthuysen, L. Moons, E. Storkebaum, H. Beck, D. Nuyens, K. Brusselmans, J. Van Dorpe, P. Hellings, M. Gorselink, S. Heymans, G. Theilmeier, M. Dewerchin, V. Laidenbach, P. Vermylen, H. Raat, T. Acker, V. Vleminckx, L. Van Den Bosch, N. Cashman, H. Fujisawa, M.R. Drost, R. Sciot, F. Bruyninckx, D.J. Hicklin, C. Ince, P. Gressens, F. Lupu, K.H. Plate, W. Robberecht, J.M. Herbert, D. Collen, P. Carmeliet, Deletion of the hypoxia-response element in the vascular endothelial growth factor promoter causes motor neuron degeneration, *Nat. Genet.* 28 (2001) 131–138.
- [60] D. Lambrechts, E. Storkebaum, M. Morimoto, J. Del-Favero, F. Desmet, S.L. Marklund, S. Wyns, V. Thijs, J. Andersson, I. van Marion, A. Al-Chalabi, S. Bornes, R. Musson, V. Hansen, L. Beckman, R. Adolfsson, H.S. Pall, H. Prats, S. Vermeire, P. Rutgeerts, S. Katayama, T. Awata, N. Leigh, L. Lang-Lazdunski, M. Dewerchin, C. Shaw, L. Moons, R. Vlietinck, K.E. Morrison, W. Robberecht, C. Van Broeckhoven, D. Collen, P.M. Andersen, P. Carmeliet, VEGF is a modifier of amyotrophic lateral sclerosis in mice and humans and protects motoneurons against ischemic death, *Nat. Genet.* 34 (2003) 383–394.
- [61] P.W. Van Vught, N.A. Sutedja, J.H. Veldink, B.P. Koeleman, G.J. Groeneveld, C. Wijmenga, B.M. Uitendaele, J.M. de Jong, F. Baas, J.H. Wokke, L.H. Van den Berg, Lack of association between VEGF polymorphisms and ALS in a Dutch population, *Neurology* 65 (2005) 1643–1645.
- [62] Y. Zhang, H. Zhang, Y. Fu, H. Song, L. Wang, J. Zhang, D. Fan, VEGF C2578A polymorphism does not contribute to amyotrophic lateral sclerosis susceptibility in sporadic Chinese patients, *Amyotroph. Lateral. Scler. Other Mot. Neuron Disord.* 7 (2006) 119–122.
- [63] A. Brockington, J. Kirby, D. Eggitt, E. Schofield, C. Morris, C.E. Lewis, P.G. Ince, P.J. Shaw, Screening of the regulatory and coding regions of vascular endothelial growth factor in amyotrophic lateral sclerosis, *Neurogenetics* 6 (2005) 101–104.
- [64] R. Fernandez-Santiago, M. Sharma, J.C. Mueller, H. Gohlke, T. Illig, J. Anneser, C. Munch, A. Ludolph, C. Kamm, T. Gasser, Possible gender-dependent association of vascular endothelial growth factor (VEGF) gene and ALS, *Neurology* 66 (2006) 1929–1931.
- [65] M.J. Greenway, P.M. Andersen, C. Russ, S. Ennis, S. Cashman, C. Donaghy, V. Patterson, R. Swigler, D. Kieran, J. Prehn, K.E. Morrison, A. Green, K.R. Acharya, R.H. Brown, O. Hardiman, ANG mutations

- segregate with familial and 'sporadic' amyotrophic lateral sclerosis, *Nat. Genet.* (2006).
- [66] M.B. Delisle, S. Carpenter, Neurofibrillary axonal swellings and amyotrophic lateral sclerosis, *J. Neurol. Sci.* 63 (1984) 241–250.
- [67] S. Matsumoto, H. Kusaka, N. Murakami, Y. Hashizume, H. Okazaki, A. Hirano, Basophilic inclusions in sporadic juvenile amyotrophic lateral sclerosis: an immunocytochemical and ultrastructural study, *Acta Neuropathol. (Berl.)* 83 (1992) 579–583.
- [68] H. Mizusawa, S. Matsumoto, S.H. Yen, A. Hirano, R.R. Rojas-Corona, H. Donnenfeld, Focal accumulation of phosphorylated neurofilaments within anterior horn cell in familial amyotrophic lateral sclerosis, *Acta Neuropathol. (Berl.)* 79 (1989) 37–43.
- [69] D. Troost, P.A. Sillevs Smitt, J.M. de Jong, D.F. Swaab, Neurofilament and glial alterations in the cerebral cortex in amyotrophic lateral sclerosis, *Acta Neuropathol. (Berl.)* 84 (1992) 664–673.
- [70] F. Cote, J.F. Collard, J.P. Julien, Progressive neuronopathy in transgenic mice expressing the human neurofilament heavy gene: a mouse model of amyotrophic lateral sclerosis, *Cell* 73 (1993) 35–46.
- [71] Z. Xu, L.C. Cork, J.W. Griffin, D.W. Cleveland, Increased expression of neurofilament subunit NF-L produces morphological alterations that resemble the pathology of human motor neuron disease, *Cell* 73 (1993) 23–33.
- [72] T.L. Williamson, L.I. Bruijn, Q. Zhu, K.L. Anderson, S.D. Anderson, J.P. Julien, D.W. Cleveland, Absence of neurofilaments reduces the selective vulnerability of motor neurons and slows disease caused by a familial amyotrophic lateral sclerosis-linked superoxide dismutase 1 mutant, *Proc. Natl. Acad. Sci. U. S. A.* 95 (1998) 9631–9636.
- [73] S. Couillard-Despres, Q. Zhu, P.C. Wong, D.L. Price, D.W. Cleveland, J.P. Julien, Protective effect of neurofilament heavy gene overexpression in motor neuron disease induced by mutant superoxide dismutase, *Proc. Natl. Acad. Sci. U. S. A.* 95 (1998) 9626–9630.
- [74] J. Kong, Z. Xu, Overexpression of neurofilament subunit NF-L and NF-H extends survival of a mouse model for amyotrophic lateral sclerosis, *Neurosci. Lett.* 281 (2000) 72–74.
- [75] S. Couillard-Despres, J. Meier, J.P. Julien, Extra axonal neurofilaments do not exacerbate disease caused by mutant Cu,Zn superoxide dismutase, *Neurobiol. Dis.* 7 (2000) 462–470.
- [76] D.A. Figlewicz, G.A. Rouleau, A. Krizus, J.P. Julien, Polymorphism in the multi-phosphorylation domain of the human neurofilament heavy-subunit-encoding gene, *Gene* 132 (1993) 297–300.
- [77] D.A. Figlewicz, A. Krizus, M.G. Martinoli, V. Meininger, M. Dib, G. A. Rouleau, J.P. Julien, Variants of the heavy neurofilament subunit are associated with the development of amyotrophic lateral sclerosis, *Hum. Mol. Genet.* 3 (1994) 1757–1761.
- [78] K. Rooke, D.A. Figlewicz, F.Y. Han, G.A. Rouleau, Analysis of the KSP repeat of the neurofilament heavy subunit in familial amyotrophic lateral sclerosis, *Neurology* 46 (1996) 789–790.
- [79] J.D. Vechio, L.I. Bruijn, Z. Xu, R.H. Brown Jr., D.W. Cleveland, Sequence variants in human neurofilament proteins: absence of linkage to familial amyotrophic lateral sclerosis, *Ann. Neurol.* 40 (1996) 603–610.
- [80] A. Al-Chalabi, P.M. Andersen, P. Nilsson, B. Chioza, J.L. Andersson, C. Russ, C.E. Shaw, J.F. Powell, P.N. Leigh, Deletions of the heavy neurofilament subunit tail in amyotrophic lateral sclerosis, *Hum. Mol. Genet.* 8 (1999) 157–164.
- [81] J. Tomkins, P. Usher, J.Y. Slade, P.G. Ince, A. Curtis, K. Bushby, P.J. Shaw, Novel insertion in the KSP region of the neurofilament heavy gene in amyotrophic lateral sclerosis (ALS), *NeuroReport* 9 (1998) 3967–3970.
- [82] M.L. Garcia, A.B. Singleton, D. Hernandez, C.M. Ward, C. Evey, P.A. Sapp, J. Hardy, R.H. Brown Jr., D.W. Cleveland, Mutations in neurofilament genes are not a significant primary cause of non-SOD1-mediated amyotrophic lateral sclerosis, *Neurobiol. Dis.* 21 (2006) 102–109.
- [83] D.M. Georgiou, J. Zidar, M. Korosec, L.T. Middleton, T. Kyriakides, K. Christodoulou, A novel NF-L mutation Pro22Ser is associated with CMT2 in a large Slovenian family, *Neurogenetics* 4 (2002) 93–96.
- [84] I.V. Mersyanova, A.V. Perepelov, A.V. Polyakov, V.F. Sitnikov, E.L. Dadali, R.B. Oparin, A.N. Petrin, O.V. Evgrafov, A new variant of Charcot–Marie–Tooth disease type 2 is probably the result of a mutation in the neurofilament-light gene, *Am. J. Hum. Genet.* 67 (2000) 37–46.
- [85] M.M. Portier, B. de Nechaud, F. Gros, Peripherin, a new member of the intermediate filament protein family, *Dev. Neurosci.* 6 (1983) 335–344.
- [86] F. Gros-Louis, R. Lariviere, G. Gowing, S. Laurent, W. Camu, J.P. Bouchard, V. Meininger, G.A. Rouleau, J.P. Julien, A frameshift deletion in peripherin gene associated with amyotrophic lateral sclerosis, *J. Biol. Chem.* 279 (2004) 45951–45956.
- [87] C.L. Leung, C.Z. He, P. Kaufmann, S.S. Chin, A. Naini, R.K. Liem, H. Mitsumoto, A.P. Hays, A pathogenic peripherin gene mutation in a patient with amyotrophic lateral sclerosis, *Brain Pathol.* 14 (2004) 290–296.
- [88] T. Murakami, I. Nagano, T. Hayashi, Y. Manabe, M. Shoji, Y. Setoguchi, K. Abe, Impaired retrograde axonal transport of adenovirus-mediated *E. coli* LacZ gene in the mice carrying mutant SOD1 gene, *Neurosci. Lett.* 308 (2001) 149–152.
- [89] A. Harada, Y. Takei, Y. Kanai, Y. Tanaka, S. Nonaka, N. Hirokawa, Golgi vesiculation and lysosome dispersion in cells lacking cytoplasmic dynein, *J. Cell Biol.* 141 (1998) 51–59.
- [90] M. Hafezparast, R. Klocke, C. Ruhrberg, A. Marquardt, A. Ahmad-Annuar, S. Bowen, G. Lalli, A.S. Witherden, H. Hummerich, S. Nicholson, P.J. Morgan, R. Oozageer, J.V. Priestley, S. Averill, V.R. King, S. Ball, J. Peters, T. Toda, A. Yamamoto, Y. Hiraoka, M. Augustin, D. Korthaus, S. Wattler, P. Wabnitz, C. Dickneite, S. Lampel, F. Boehme, G. Peraus, A. Popp, M. Rudelius, J. Schlegel, H. Fuchs, M. Hrabe de Angelis, G. Schiavo, D.T. Shima, A.P. Russ, G. Stumm, J.E. Martin, E.M. Fisher, Mutations in dynein link motor neuron degeneration to defects in retrograde transport, *Science* 300 (2003) 808–812.
- [91] A. Ahmad-Annuar, P. Shah, M. Hafezparast, H. Hummerich, A.S. Witherden, K.E. Morrison, P.J. Shaw, J. Kirby, T.T. Warner, A. Crosby, C. Proukakakis, P. Wilkinson, R.W. Orrell, L. Bradley, J.E. Martin, E.M. Fisher, No association with common Caucasian genotypes in exons 8, 13 and 14 of the human cytoplasmic dynein heavy chain gene (DNCHC1) and familial motor neuron disorders, *Amyotroph. Lateral. Scler. Other Mot. Neuron Disord.* 4 (2003) 150–157.
- [92] L.A. Ligon, B.H. LaMonte, K.E. Wallace, N. Weber, R.G. Kalb, E.L. Holzbaur, Mutant superoxide dismutase disrupts cytoplasmic dynein in motor neurons, *NeuroReport* 16 (2005) 533–536.
- [93] B.A. Eaton, R.D. Fetter, G.W. Davis, Dynactin is necessary for synapse stabilization, *Neuron* 34 (2002) 729–741.
- [94] B.H. LaMonte, K.E. Wallace, B.A. Holloway, S.S. Shelly, J. Ascano, M. Tokito, T. Van Winkle, D.S. Howland, E.L. Holzbaur, Disruption of dynein/dynactin inhibits axonal transport in motor neurons causing late-onset progressive degeneration, *Neuron* 34 (2002) 715–727.
- [95] C. Munch, R. Sedlmeier, T. Meyer, V. Homberg, A.D. Sperfeld, A. Kurt, J. Prudlo, G. Peraus, C.O. Hanemann, G. Stumm, A.C. Ludolph, Point mutations of the p150 subunit of dynactin (DCTN1) gene in ALS, *Neurology* 63 (2004) 724–726.
- [96] I. Puls, C. Jonnakuty, B.H. LaMonte, E.L. Holzbaur, M. Tokito, E. Mann, M.K. Floeter, K. Bidus, D. Drayna, S.J. Oh, R.H. Brown Jr., C.L. Ludlow, K.H. Fischbeck, Mutant dynactin in motor neuron disease, *Nat. Genet.* 33 (2003) 455–456.
- [97] C. Munch, A. Rosenbohm, A.D. Sperfeld, I. Uttner, S. Reske, B.J. Krause, R. Sedlmeier, T. Meyer, C.O. Hanemann, G. Stumm, A.C. Ludolph, Heterozygous R1101K mutation of the DCTN1 gene in a family with ALS and FTD, *Ann. Neurol.* 58 (2005) 777–780.
- [98] P. Corcia, V. Mayeux-Portas, J. Khoris, B. de Toffol, A. Autret, J.P. Muh, W. Camu, C. Andres, Abnormal SMN1 gene copy number is a susceptibility factor for amyotrophic lateral sclerosis, *Ann. Neurol.* 51 (2002) 243–246.
- [99] M. Jackson, K.E. Morrison, A. Al-Chalabi, M. Bakker, P.N. Leigh, Analysis of chromosome 5q13 genes in amyotrophic lateral sclerosis: homozygous NAIP deletion in a sporadic case, *Ann. Neurol.* 39 (1996) 796–800.
- [100] B. Moulard, F. Salachas, B. Chassande, V. Briolotti, V. Meininger, A. Malafose, W. Camu, Association between centromeric deletions of the SMN gene and sporadic adult-onset lower motor neuron disease, *Ann. Neurol.* 43 (1998) 640–644.

- [101] R.W. Orrell, J.J. Habgood, J.S. de Belleruche, R.J. Lane, The relationship of spinal muscular atrophy to motor neuron disease: investigation of SMN and NAIP gene deletions in sporadic and familial ALS, *J. Neurol. Sci.* 145 (1997) 55–61.
- [102] J.S. Parboosingh, V. Meininger, D. McKenna-Yasek, R.H. Brown Jr., G.A. Rouleau, Deletions causing spinal muscular atrophy do not predispose to amyotrophic lateral sclerosis, *Arch. Neurol.* 56 (1999) 710–712.
- [103] J. Gamez, M.J. Barcelo, X. Munoz, F. Carmona, I. Cusco, M. Baiget, C. Cervera, E.F. Tizzano, Survival and respiratory decline are not related to homozygous SMN2 deletions in ALS patients, *Neurology* 59 (2002) 1456–1460.
- [104] J.H. Veldink, L.H. van den Berg, J.M. Cobben, R.P. Stulp, J.M. De Jong, O.J. Vogels, F. Baas, J.H. Wokke, H. Scheffer, Homozygous deletion of the survival motor neuron 2 gene is a prognostic factor in sporadic ALS, *Neurology* 56 (2001) 749–752.
- [105] J.H. Veldink, S. Kalmijn, A.H. Van der Hout, H.H. Lemmink, G.J. Groeneveld, C. Lummen, H. Scheffer, J.H. Wokke, L.H. Van den Berg, SMN genotypes producing less SMN protein increase susceptibility to and severity of sporadic ALS, *Neurology* 65 (2005) 820–825.
- [106] C.B. Kunst, L. Messer, J. Gordon, J. Haines, D. Patterson, Genetic mapping of a mouse modifier gene that can prevent ALS onset, *Genomics* 70 (2000) 181–189.
- [107] F. Kamel, D.M. Umbach, T.A. Lehman, L.P. Park, T.L. Munsat, J.M. Shefner, D.P. Sandler, H. Hu, J.A. Taylor, Amyotrophic lateral sclerosis, lead, and genetic susceptibility: polymorphisms in the delta-aminolevulinic acid dehydratase and vitamin D receptor genes, *Environ. Health Perspect.* 111 (2003) 1335–1339.
- [108] F. Kamel, D.M. Umbach, T.L. Munsat, J.M. Shefner, H. Hu, D.P. Sandler, Lead exposure and amyotrophic lateral sclerosis, *Epidemiology* 13 (2002) 311–319.
- [109] C.K. Hand, R.S. Devon, F. Gros-Louis, D. Rochefort, J. Khoris, V. Meininger, J.P. Bouchard, W. Camu, M.R. Hayden, G.A. Rouleau, Mutation screening of the ALS2 gene in sporadic and familial amyotrophic lateral sclerosis, *Arch. Neurol.* 60 (2003) 1768–1771.
- [110] S. Hadano, C.K. Hand, H. Osuga, Y. Yanagisawa, A. Otomo, R.S. Devon, N. Miyamoto, J. Showguchi-Miyata, Y. Okada, R. Singaraja, D.A. Figlewicz, T. Kwiatkowski, B.A. Hosler, T. Sagie, J. Skaug, J. Nasir, R.H. Brown Jr., S.W. Scherer, G.A. Rouleau, M.R. Hayden, J.E. Ikeda, A gene encoding a putative GTPase regulator is mutated in familial amyotrophic lateral sclerosis 2, *Nat. Genet.* 29 (2001) 166–173.
- [111] Y. Yang, A. Hentati, H.X. Deng, O. Dabbagh, T. Sasaki, M. Hirano, W.Y. Hung, K. Ouahchi, J. Yan, A.C. Azim, N. Cole, G. Gascon, A. Yagmour, M. Ben-Hamida, M. Pericak-Vance, F. Hentati, T. Siddique, The gene encoding alsin, a protein with three guanine-nucleotide exchange factor domains, is mutated in a form of recessive amyotrophic lateral sclerosis, *Nat. Genet.* 29 (2001) 160–165.
- [112] B.A. Hosler, P.C. Sapp, R. Berger, G. O'Neill, K. Bejaoui, M.B. Hamida, F. Hentati, W. Chin, D. McKenna-Yasek, J.L. Haines, D. Patterson, H.R. Horvitz, R.H. Brown Jr., C.B. Day, Refined mapping and characterization of the recessive familial amyotrophic lateral sclerosis locus (ALS2) on chromosome 2q33, *Neurogenetics* 2 (1998) 34–42.
- [113] A. Hentati, K. Bejaoui, M.A. Pericak-Vance, F. Hentati, M.C. Speer, W.Y. Hung, D.A. Figlewicz, J. Haines, J. Rimmler, C. Ben Hamida, et al., Linkage of recessive familial amyotrophic lateral sclerosis to chromosome 2q33–q35, *Nat. Genet.* 7 (1994) 425–428.
- [114] M.J. Greenway, M.D. Alexander, S. Ennis, B.J. Traynor, B. Corr, E. Frost, A. Green, O. Hardiman, A novel candidate region for ALS on chromosome 14q11.2, *Neurology* 63 (2004) 1936–1938.
- [115] C. Hayward, S. Colville, R.J. Swingle, D.J. Brock, Molecular genetic analysis of the APEX nuclease gene in amyotrophic lateral sclerosis, *Neurology* 52 (1999) 1899–1901.
- [116] O. Garofalo, D.A. Figlewicz, P.N. Leigh, J.F. Powell, V. Meininger, M. Dib, G.A. Rouleau, Androgen receptor gene polymorphisms in amyotrophic lateral sclerosis, *Neuromuscul. Disord.* 3 (1993) 195–199.
- [117] A.N. Silahatoglu, K. Brondum-Nielsen, O. Gredal, L. Werdelin, M. Panas, M.B. Petersen, N. Tommerup, Z. Tumer, Human CCS gene: genomic organization and exclusion as a candidate for amyotrophic lateral sclerosis (ALS), *BMC Genet.* 3 (2002) 5.
- [118] T.M. DeChiara, R. Vejsada, W.T. Poueymirou, A. Acheson, C. Suri, J.C. Conover, B. Friedman, J. McClain, L. Pan, N. Stahl, et al., Mice lacking the CNTF receptor, unlike mice lacking CNTF, exhibit profound motor neuron deficits at birth, *Cell* 83 (1995) 313–322.
- [119] R. Giess, B. Holtmann, M. Braga, T. Grimm, B. Muller-Mysok, K.V. Toyka, M. Sendtner, Early onset of severe familial amyotrophic lateral sclerosis with a SOD-1 mutation: potential impact of CNTF as a candidate modifier gene, *Am. J. Hum. Genet.* 70 (2002) 1277–1286.
- [120] D.J. Nicholl, P. Bennett, L. Hiller, V. Bonifati, N. Vanacore, G. Fabbri, R. Marconi, C. Colosimo, P. Lamberti, F. Stocchi, U. Bonuccelli, P. Vieregge, D.B. Ramsden, G. Meco, A.C. Williams, A study of five candidate genes in Parkinson's disease and related neurodegenerative disorders. European Study Group on Atypical Parkinsonism, *Neurology* 53 (1999) 1415–1421.
- [121] M.A. Siddons, S.M. Pickering-Brown, D.M. Mann, F. Owen, P.N. Cooper, Debrisoquine hydroxylase gene polymorphism frequencies in patients with amyotrophic lateral sclerosis, *Neurosci. Lett.* 208 (1996) 65–68.
- [122] J.M. Flowers, J.F. Powell, P.N. Leigh, P. Andersen, C.E. Shaw, Intron 7 retention and exon 9 skipping EAAT2 mRNA variants are not associated with amyotrophic lateral sclerosis, *Ann. Neurol.* 49 (2001) 643–649.
- [123] L.S. Honig, D.D. Chambliss, E.H. Bigio, S.L. Carroll, J.L. Elliott, Glutamate transporter EAAT2 splice variants occur not only in ALS, but also in AD and controls, *Neurology* 55 (2000) 1082–1088.
- [124] M. Jackson, G. Steers, P.N. Leigh, K.E. Morrison, Polymorphisms in the glutamate transporter gene EAAT2 in European ALS patients, *J. Neurol.* 246 (1999) 1140–1144.
- [125] T. Meyer, A. Fromm, C. Munch, B. Schwalenstocker, A.E. Fray, P.G. Ince, S. Stamm, G. Gron, A.C. Ludolph, P.J. Shaw, The RNA of the glutamate transporter EAAT2 is variably spliced in amyotrophic lateral sclerosis and normal individuals, *J. Neurol. Sci.* 170 (1999) 45–50.
- [126] T. Meyer, C. Munch, H. Volkel, P. Booms, A.C. Ludolph, The EAAT2 (GLT-1) gene in motor neuron disease: absence of mutations in amyotrophic lateral sclerosis and a point mutation in patients with hereditary spastic paraplegia, *J. Neurol., Neurosurg. Psychiatry* 65 (1998) 594–596.
- [127] M. Aoki, C.L. Lin, J.D. Rothstein, B.A. Geller, B.A. Hosler, T.L. Munsat, H.R. Horvitz, R.H. Brown Jr., Mutations in the glutamate transporter EAAT2 gene do not cause abnormal EAAT2 transcripts in amyotrophic lateral sclerosis, *Ann. Neurol.* 43 (1998) 645–653.
- [128] C.L. Lin, L.A. Bristol, L. Jin, M. Dykes-Hoberg, T. Crawford, L. Clawson, J.D. Rothstein, Aberrant RNA processing in a neurodegenerative disease: the cause for absent EAAT2, a glutamate transporter, in amyotrophic lateral sclerosis, *Neuron* 20 (1998) 589–602.
- [129] V.E. Drory, M. Birnbaum, L. Peleg, B. Goldman, A.D. Korczyn, Hexosaminidase A deficiency is an uncommon cause of a syndrome mimicking amyotrophic lateral sclerosis, *Muscle Nerve* 28 (2003) 109–112.
- [130] X.S. Wang, S. Lee, Z. Simmons, P. Boyer, K. Scott, W. Liu, J. Connor, Increased incidence of the Hfe mutation in amyotrophic lateral sclerosis and related cellular consequences, *J. Neurol. Sci.* 227 (2004) 27–33.
- [131] A.A. Yen, E.P. Simpson, J.S. Henkel, D.R. Beers, S.H. Appel, HFE mutations are not strongly associated with sporadic ALS, *Neurology* 62 (2004) 1611–1612.
- [132] R. Giess, M. Beck, R. Goetz, R.M. Nitsch, K.V. Toyka, M. Sendtner, Potential role of LIF as a modifier gene in the pathogenesis of amyotrophic lateral sclerosis, *Neurology* 54 (2000) 1003–1005.
- [133] M.A. Meyer, N.T. Potter, Sporadic ALS and chromosome 22: evidence for a possible neurofilament gene defect, *Muscle Nerve* 18 (1995) 536–539.
- [134] B.A. Chioza, A. Ujfalusy, K. Csiszar, P.N. Leigh, J.F. Powell, A. Radunovic, Mutations in the lysyl oxidase gene are not associated with amyotrophic lateral sclerosis, *Amyotroph. Lateral. Scler. Other Mot. Neuron Disord.* 2 (2001) 93–97.
- [135] S. Orru, V. Mascia, M. Casula, E. Giurelli, A. Loizadda, C. Carcassi, M. Giagghedu, L. Contu, Association of monoamine oxidase B alleles with age at onset in amyotrophic lateral sclerosis, *Neuromuscul. Disord.* 9 (1999) 593–597.

- [136] A. Kowalska, M. Konagaya, M. Sakai, Y. Hashizume, T. Tabira, Familial amyotrophic lateral sclerosis and parkinsonism–dementia complex-tauopathy without mutations in the tau gene? *Folia Neuropathol.* 41 (2003) 59–64.
- [137] P. Poorkaj, D. Tsuang, E. Wijsman, E. Steinbart, R.M. Garruto, U.K. Craig, N.H. Chapman, L. Anderson, T.D. Bird, C.C. Plato, D.P. Perl, W. Weiderholt, D. Galasko, G.D. Schellenberg, TAU as a susceptibility gene for amyotrophic lateral sclerosis–parkinsonism dementia complex of Guam, *Arch. Neurol.* 58 (2001) 1871–1878.
- [138] G.K. Dhaliwal, R.P. Grewal, Mitochondrial DNA deletion mutation levels are elevated in ALS brains, *NeuroReport* 11 (2000) 2507–2509.
- [139] C.D. Gajewski, M.T. Lin, M.E. Cudkowicz, M.F. Beal, G. Manfredi, Mitochondrial DNA from platelets of sporadic ALS patients restores normal respiratory functions in rho(0) cells, *Exp. Neurol.* 179 (2003) 229–235.
- [140] C. Mawrin, E. Kirches, K. Dietzmann, Single-cell analysis of mtDNA in amyotrophic lateral sclerosis: towards the characterization of individual neurons in neurodegenerative disorders, *Pathol. Res. Pract.* 199 (2003) 415–418.
- [141] C. Mawrin, E. Kirches, G. Krause, F.R. Wiedemann, C.K. Vorwerk, B. Bogerts, H.U. Schildhaus, K. Dietzmann, R. Schneider-Stock, Single-cell analysis of mtDNA deletion levels in sporadic amyotrophic lateral sclerosis, *NeuroReport* 15 (2004) 939–943.
- [142] L.S. Ro, S.L. Lai, C.M. Chen, S.T. Chen, Deleted 4977-bp mitochondrial DNA mutation is associated with sporadic amyotrophic lateral sclerosis: a hospital-based case-control study, *Muscle Nerve* 28 (2003) 737–743.
- [143] R.H. Swerdlow, J.K. Parks, D.S. Cassarino, P.A. Trimmer, S.W. Miller, D.J. Maguire, J.P. Sheehan, R.S. Maguire, G. Pattee, V.C. Juel, L.H. Phillips, J.B. Tuttle, J.P. Bennett Jr., R.E. Davis, W.D. Parker Jr., Mitochondria in sporadic amyotrophic lateral sclerosis, *Exp. Neurol.* 153 (1998) 135–142.
- [144] F.R. Wiedemann, G. Manfredi, C. Mawrin, M.F. Beal, E.A. Schon, Mitochondrial DNA and respiratory chain function in spinal cords of ALS patients, *J. Neurochem.* 80 (2002) 616–625.
- [145] F.H. Lin, R. Lin, H.M. Wisniewski, Y.W. Hwang, I. Grundke-Iqbal, G. Healy-Louie, K. Iqbal, Detection of point mutations in codon 331 of mitochondrial NADH dehydrogenase subunit 2 in Alzheimer's brains, *Biochem. Biophys. Res. Commun.* 182 (1992) 238–246.
- [146] J.P. Julien, F. Cote, J.F. Collard, Mice overexpressing the human neurofilament heavy gene as a model of ALS, *Neurobiol. Aging* 16 (1995) 487–490 (discussion 490–2).
- [147] M. Panas, G. Karadima, N. Kalfakis, O. Psarrou, P. Floroskoufi, A. Kladi, M.B. Petersen, D. Vassilopoulos, Genotyping of presenilin-1 polymorphism in amyotrophic lateral sclerosis, *J. Neurol.* 247 (2000) 940–942.
- [148] R. Saunderson, B. Yu, R.J. Trent, R. Pamphlett, A polymorphism in the poliovirus receptor gene differs in motor neuron disease, *NeuroReport* 15 (2004) 383–386.
- [149] Y.Z. Chen, C.L. Bennett, H.M. Huynh, I.P. Blair, I. Puls, J. Irobi, I. Dierick, A. Abel, M.L. Kennerson, B.A. Rabin, G.A. Nicholson, M. Auer-Grumbach, K. Wagner, P. De Jonghe, J.W. Griffin, K.H. Fischbeck, V. Timmerman, D.R. Cornblath, P.F. Chance, DNA/RNA helicase gene mutations in a form of juvenile amyotrophic lateral sclerosis (ALS4), *Am. J. Hum. Genet.* 74 (2004) 1128–1135.
- [150] P. Corcia, J. Khoris, P. Couratier, V. Mayeux-Portas, E. Bieth, B. De Toffol, A. Autret, J.P. Muh, C. Andres, W. Camu, SMN1 gene study in three families in which ALS and spinal muscular atrophy co-exist, *Neurology* 59 (2002) 1464–1466.
- [151] C.J. McDermott, D. Roberts, J. Tomkins, K.M. Bushby, P.J. Shaw, Spastin and paraplegin gene analysis in selected cases of motor neurone disease (MND), *Amyotroph. Lateral. Scler. Other Mot. Neuron Disord.* 4 (2003) 96–99.
- [152] T. Meyer, A. Schwan, J.S. Dullinger, J. Brocke, K.T. Hoffmann, C.H. Nolte, A. Hopt, U. Kopp, P. Andersen, J.T. Epplen, P. Linke, Early-onset ALS with long-term survival associated with spastin gene mutation, *Neurology* 65 (2005) 141–143.
- [153] J.M. Flowers, P.N. Leigh, A.M. Davies, N.N. Ninkina, V.L. Buchman, J. Vaughan, N.W. Wood, J.F. Powell, Mutations in the gene encoding human peryn are not associated with amyotrophic lateral sclerosis or familial Parkinson's disease, *Neurosci. Lett.* 274 (1999) 21–24.
- [154] J. Tomkins, S.J. Banner, C.J. McDermott, P.J. Shaw, Mutation screening of manganese superoxide dismutase in amyotrophic lateral sclerosis, *NeuroReport* 12 (2001) 2319–2322.
- [155] A.L. Nishimura, M. Mitne-Neto, H.C. Silva, A. Richieri-Costa, S. Middleton, D. Cascio, F. Kok, J.R. Oliveira, T. Gillingwater, J. Webb, P. Skehel, M. Zatz, A mutation in the vesicle-trafficking protein VAPB causes late-onset spinal muscular atrophy and amyotrophic lateral sclerosis, *Am. J. Hum. Genet.* 75 (2004) 822–831.
- [156] P.D. Terry, F. Kamel, D.M. Umbach, T.A. Lehman, H. Hu, D.P. Sandler, J.A. Taylor, VEGF promoter haplotype and amyotrophic lateral sclerosis (ALS), *J. Neurogenet.* 18 (2004) 429–434.